

Chapter 10

Spatial topography of the basal forebrain cholinergic projections: Organization and vulnerability to degeneration

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Abstract

The basal forebrain (BF) cholinergic system constitutes a heterogeneous cluster of large projection neurons that innervate the entire cortical mantle and amygdala. Cholinergic neuromodulation plays a critical role in regulating cognition and behavior, as well as maintenance of cellular homeostasis. Decades of postmortem histology research have demonstrated that the BF cholinergic neurons are selectively vulnerable to aging and age-related neuropathology in degenerative diseases such as Alzheimer's and Parkinson's diseases. Emerging evidence from in vivo neuroimaging research, which permits longitudinal tracking of at-risk individuals, indicates that cholinergic neurodegeneration might play an earlier and more pivotal role in these diseases than was previously appreciated. Despite these advances, our understanding of the organization and functions of the BF cholinergic system mostly derives from nonhuman animal research. In this chapter, we begin with a review of the topographical organization of the BF cholinergic system in rodent and nonhuman primate models. We then discuss basic and clinical neuroscience research in humans, which has started to translate and extend the nonhuman animal research using novel noninvasive neuroimaging techniques. We focus on converging evidence indicating that the selective vulnerability of cholinergic neurons in Alzheimer's and Parkinson's diseases is expressed along a rostral-caudal topography in the BF. We close with a discussion of why this topography of vulnerability in the BF may occur and why it is relevant to the clinician.

INTRODUCTION

Basic and clinical neuroscience research on the basal forebrain (BF) cholinergic system has experienced somewhat of a renaissance in the past 15 years. Driven by advances in cell labeling techniques, electrophysiology and neuroimaging, our understanding of the organization and function of the BF cholinergic system and its primary neurochemical, acetylcholine, is coming into sharper focus. While acetylcholine was previously thought to operate in a single mode of regulating general states of

alertness, characterized by global neuromodulation of cortical excitability, nonhuman animal research has demonstrated that this neurochemical is capable of modulating cortical activity at multiple spatial and temporal scales. In addition to the global excitation mode, acetylcholine can also excite or inhibit local cortical ensembles in time scales associated with specific cognitive and behavioral events, a mode which resembles wired neurotransmission (Sarter et al., 2009; Picciotto et al., 2012; Muñoz and Rudy, 2014; Ballinger et al., 2016; Obermayer et al., 2017; Schmitz and Duncan, 2018;

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Zaborszky et al., 2018; Sarter and Lustig, 2020). Parallelizing the recent advances in basic research, there is renewed clinical research interest in the BF system as both a preclinical biomarker of neurodegenerative disease and as a target of therapeutic intervention (Hampel et al., 2018). Building on an extensive history of postmortem histology work dating from the 1980s, recent in vivo clinical neuroscience research on Alzheimer's and Parkinson's disease (AD and PD), has demonstrated that degeneration of the BF system is closely linked to cognitive impairments in both early AD (Grothe et al., 2010, 2012; Cantero et al., 2017; Scheef et al., 2019; Nemy et al., 2020) and PD (Ray et al., 2018; Schulz et al., 2018; Pereira et al., 2020). Emerging evidence suggests that structural degeneration of BF neurons occurs much earlier in these conditions than was previously thought, and may in fact precede, predict, and even potentiate cortical degeneration (Schmitz and Spreng, 2016; Schmitz et al., 2018; Fernandez-Cabello et al., 2020).

In the following, three sections of this chapter, we begin with a review of the first attempts to standardize the anatomy of the BF cholinergic system using nonhuman primate and rodent models. We then discuss more recent work in these nonhuman animal models, which has shed new light on the organizing principles of the cortical cholinergic projections, namely their cortical topography. In the second section of this chapter, we discuss recent in vivo neuroimaging work in humans, which has started to translate and extend the work from nonhuman animal models. This work shows that structural magnetic resonance imaging, functional MRI (fMRI), and vesicular acetylcholine transporter positron emission tomography (VACHT PET) are promising techniques for mapping the topography of the cholinergic projection system and deepening our understanding of its role in cognition. In the third and final section of this chapter, we discuss emerging evidence in human neuroimaging work indicating that the vulnerability of cholinergic BF neurons to aging and neurodegenerative disease also follows a subregional topography.

TOPOGRAPHIC PROJECTIONS OF THE BF IN RODENTS AND NONHUMAN PRIMATES

The nonhuman primate BF comprises a set of large hyperchromic neurons, also referred as magnocellular neurons, which provide the major cholinergic innervation to the cerebral cortex (Hedreen et al., 1984; Saper and Chelimsky, 1984; Mesulam and Geula, 1988). Early histology work showed that the majority of these magnocellular neurons project to large and discrete cortical regions, such as frontal, cingulate, and temporal cortices (Jones et al., 1976; Bigl et al., 1982; Pearson et al., 1983).

More recent histology work has shown that this projection pattern reflects a topography of BF inputs and outputs (Zaborszky et al., 2015; Gielow and Zaborszky, 2017). The BF cholinergic neurons located more rostrally in the medial septum and the nucleus of the diagonal band of Broca project to the hippocampus, entorhinal cortex, olfactory bulb, and cingulate cortices (Mesulam and Van Hoesen, 1976; Pearson et al., 1983; Mesulam et al., 1983a; Zaborszky et al., 1986; Woolf, 1991; Kondo and Zaborszky, 2016). The BF cholinergic neurons located more caudally in the subcommissural and sublenticular substantia innominata, often referred to as the nucleus basalis of Meynert, project throughout the entire neocortex (Jones et al., 1976; Pearson et al., 1983; Mesulam et al., 1983a; Woolf, 1991) and to the amygdala (Carlsen et al., 1985).

In the early 1980s, Mesulam et al. introduced a "Ch" nomenclature that delineates Cholinergic neurons based on their topography in the BF and their cortical targets in rodents (Mesulam et al., 1983b) and nonhuman primates (Mesulam et al., 1983a). Cholinergic cells in the medial septum and diagonal band of Broca that project to the hippocampus were designated as Ch1 and Ch2, respectively. The Ch3 group was used to refer a band of fusiform neurons that are close to the ventral surface of the brain in the subcommissural area and project to the olfactory bulb (Mesulam and Geula, 1988; Mufson et al., 1989). Those cholinergic cells projecting to the major part of the neocortex and amygdala were termed as Ch4.

Together, these early axonal tracing studies implied a rostro-caudal spatial topography of BF cholinergic innervation into the cortex and amygdala. More recent studies have shed further light on the anatomical significance of this topographic organization. Using retrograde tracers deposited into multiple frontal and posterior cortical areas of the rat brain, Zaborszky et al. (2015) demonstrated that labeled cells in the BF exhibit a complex topographic organization consisting of segregated and overlapping pools of projection neurons. The extent of this overlap seems to depend on the degree of connectivity between their cortical targets (Fig. 10.1). For example, BF cell populations projecting to cortical areas representing different body parts, such as S1 whisker and S1 hindlimb, have very low BF spatial overlap. In contrast, BF cell populations projecting to the primary somatosensory S1 and the corresponding primary motor M1 regions tend to have higher BF spatial overlap. This principle seems to extend to the level of individual BF neurons as well. Axonal branches from individual BF neurons innervate multiple cortical regions that are functionally related to one another, such as subregions of the prefrontal cortex (Chandler and Waterhouse, 2012; Chandler et al., 2013; Bloem et al., 2014; Chavez and Zaborszky, 2017). However, the branches of individual

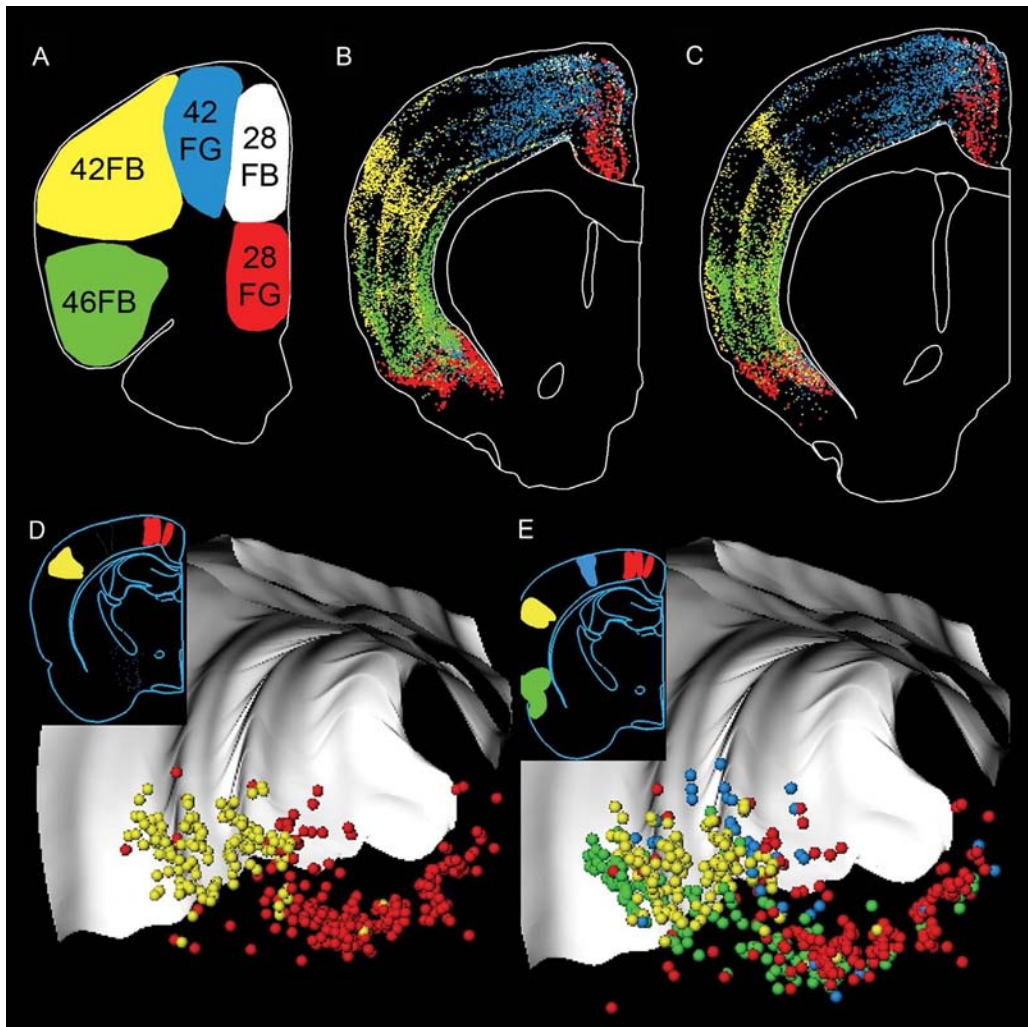


Fig. 10.1. A 3D rendering of the distribution of retrogradely labeled cells in the rat brain revealing the topographical organization of cortico-cortical (A–C) and basal forebrain (BF)-cortical (D, E) projections. When retrograde tracers are injected into different cortical locations (A), the organization of the cells in the posterior cortical areas that project to the frontal cortex becomes apparent (B and C: neurons are color coded according to their cortical injection site). The distribution of cortico-cortical projections has both rostro-caudal and medial-lateral organization. When retrograde tracers are injected into different cortical locations (D and E insets), the organization of the choline-o-acetyltransferase expressing (ChAT+) cholinergic cells in the BF that project to the frontal cortex becomes apparent (D, E: neurons are color coded according to their cortical injection site). The degree of overlap of BF neurons, as defined by their retrograde labeling from different locations in the frontal and posterior cortex (bottom row similarly colored clusters), is correlated with the amount of cortico-cortical connectivity of posterior cortical neurons, as defined by their retrograde labeling from different locations in frontal cortex (top row similarly colored clusters). Adapted from Zaborszky, L., Csordas, A., Mosca, K., et al., 2015a. Neurons in the basal forebrain project to the cortex in a complex topographic organization that reflects corticocortical connectivity patterns: an experimental study based on retrograde tracing and 3D reconstruction. *Cereb Cortex* 25, 118–137.

BF neurons do not appear to innervate functionally distinct regions of neocortex such as modality specific primary auditory A1 and primary visual V1 areas (Kim et al., 2016). Taken together, these results suggest a topographical organization of the BF projections at multiple spatial scales, from populations of BF neurons down to the axon branches of individual cells. This architecture appears well suited for coordinating information across the cortical hierarchy.

Despite these advances, the organizing principles of the mammalian BF cholinergic system and the functional properties embedded within its architecture are not fully worked out. There are three anatomical properties of the BF system that pose major challenges to its detailed experimental study: (1) the enormous and complex morphology of BF cholinergic neurons, (2) the multiplicity of postsynaptic cholinergic signaling pathways, and (3) the heterogeneity of noncholinergic neurons

codistributed within the BF. Nevertheless, genetically controlled techniques for cell-type specific labeling, stimulation, and receptor modeling are becoming available for studying the BF projection system at ever-increasing levels of precision (Letzkus et al., 2011; Arroyo et al., 2012; Pinto et al., 2013; Wu et al., 2014; Anacleto et al., 2015; Saunders et al., 2015; Unal et al., 2015; Xu et al., 2015; Do et al., 2016; Verhoog et al., 2016; Kuchibhotla et al., 2017; López-Hernández et al., 2017; Jing et al., 2018; Li et al., 2018; Laszlovszky et al., 2020). Although beyond the scope of this chapter, this work has demonstrated that the topography of the BF cholinergic system is expressed not only in terms of the input–output relationships of its cortical projections but also in the neurophysiological properties of cholinergic neurons across the rostral-caudal BF axis (Gielow and Zaborszky, 2017; Laszlovszky et al., 2020), the distributions of postsynaptic nicotinic and muscarinic cholinergic receptors in specific layers of the cortex (Verhoog et al., 2016), and the spatiotemporal dynamics of cortical state transitions (Pinto et al., 2013; Anacleto et al., 2015; Xu et al., 2015).

TOPOGRAPHIC PROJECTIONS OF THE BASAL FOREBRAIN IN HUMANS

Ex vivo localization of the human basal forebrain nuclei

Much less is understood about the organization of the human BF system compared to rodents and nonhuman primates. In human postmortem tissue samples, cholinergic BF neurons are often aggregated in clusters and constitute most of the large, or magnocellular, neurons (>20- μm -long axis) in this brain area. The delineation of these clusters on standard structural MRI is difficult to reproduce due to lack of specific landmarks, creating a major obstacle for in vivo neuroimaging of this structure.

However, the BF areas containing such magnocellular cell groups can be easily delineated in postmortem histological sections stained with a modified Gallyas's silver method for cell bodies (Merker, 1983). Using this approach, Zaborszky et al. (2008) identified clusters of hyperchromic magnocellular cell groups located close to the medial and ventral surfaces of the cerebral hemispheres (Fig. 10.2, left panels), including the septum,

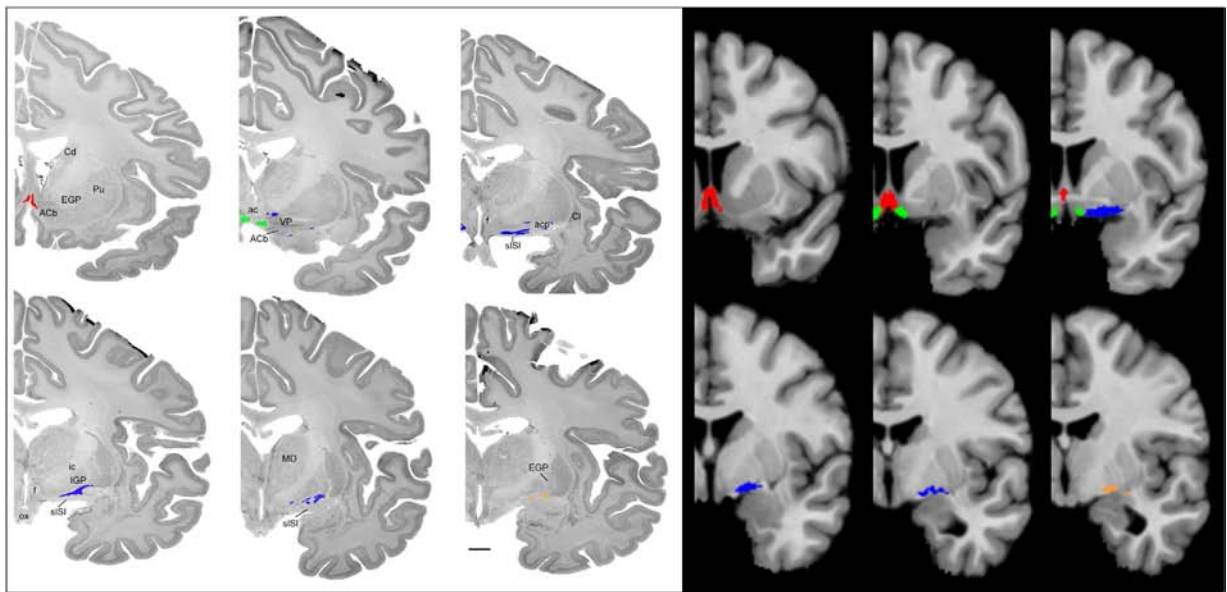


Fig. 10.2. Stereotaxic probabilistic maps of the human magnocellular basal forebrain (BF) space from Zaborszky et al. (2008) and Zaborszky et al. (2015). *Left panels:* The BF space delineated on 2D images of silver-stained histological sections of a representative brain specimen. *Right panels:* The BF space after postmortem T1-weighted MRI scanning of 10 specimens, and coregistration into a common atlas space. The colored areas delineate the maximum probability maps of the various cholinergic (Ch) compartments: Ch12 (red, medial septal area, and vertical band of Broca), Ch3 (green, horizontal band of Broca), Ch4 (blue, nucleus basalis of Meynert), and Chp (orange, nucleus of Ayala). A voxel was assigned to a BF area that showed the greatest overlap (winner take all) among the 10 examined postmortem brains or that exceeded a threshold of 40% probability (4/10 brain overlapped). As a result, maximum probability maps show a nonoverlapping representation of the Ch compartments in the Montreal Neurological Institute anatomical template space. ACb, nucleus accumbens; ac, anterior commissure; Cd, caudate nucleus; Cl, claustrum; EGP, external globus pallidus; IGP, internal globus pallidus; ic, internal capsule; ox, optic chiasm; Pu, putamen.

the horizontal and vertical limbs of the diagonal band, the basal nucleus of Meynert, and cell groups interspersed in the so-called “extended amygdala” (the area underneath the globus pallidus bridging the centromedial amygdala to the bed nucleus of the stria terminalis). Following from Mesulam’s “Ch” nomenclature for cholinergic compartments (Mesulam et al., 1983a, b), Zaborszky et al. (2008) separated these groups into medial septal and vertical band of Broca (Ch1 and Ch2, Red), the horizontal band of Broca (Ch3, Green), the nucleus basalis of Meynert (Ch4, Blue), and a posterior group behind the supraoptic nucleus, where the optic tract attaches to the internal capsule/cerebral peduncle, including the nucleus of Ayala (Ch4p, Orange).

Other atlases of the human BF Ch space have also been produced in single brain specimens using similar procedures to Zaborszky and colleagues (Teipel et al., 2005; Kilimann et al., 2014). Unlike these atlases, however, Zaborszky et al. (2008) delineated BF Ch groups in a total of 10 postmortem brains that were then digitized and coregistered into the MNI single-subject reference brain (Colin27) as a common brain atlas space, allowing the creation of a probabilistic anatomical map of the human BF compartments (Fig. 10.2, right panels). Since postmortem probabilistic maps were constructed primarily based on the presence of large neurons that are mostly cholinergic, the BF space is referred to as the BF cholinergic space. The probabilistic human BF atlas can be thresholded to infer the location of the Ch compartments at varying levels of certainty, e.g., a 50% probability threshold isolates voxels that converge on a given BF Ch compartment in 5/10 brains. However, unlike studies in nonhuman animal models (Fig. 10.1), it should be noted that the human subdivisions are defined without reference to projection targets or cell-type specific labeling.

Similar postmortem histology approaches have been taken to map the projections of the human cholinergic BF. Selden et al. (1998) used acetylcholinesterase (AChE) histochemistry to trace AChE-rich white matter fibers originating from the nucleus basalis of Meynert or Ch4 (Fig. 10.3). Consistent with a topographical organization of BF projections, they identified two primary pathways originating from Ch4 (blue). A medial pathway (green) traveled within the cingulum to supply the orbitofrontal, subcallosal, cingulate, pericingulate, and retrosplenial cortices. A lateral pathway subdivided into a capsular (red) and perisylvian (orange) component. The capsular pathway traveled within the external capsule and uncinate fasciculus to supply the putamen, amygdala, middle, and inferior temporal gyri, parahippocampal gyri, and dorsal frontoparietal neocortex. The perisylvian component traveled within the claustrum to supply the insula, superior temporal gyri, and ventral (opercular) frontoparietal neocortex. The more rostral BF Ch

compartments, including Ch1–3, were not assessed in this study. To date, the immunohistochemical mapping of the human BF cholinergic projection system remains incomplete.

In vivo MRI of the human basal forebrain nuclei and topographic projections

Taken together, the atlases of the BF Ch compartments and their projections serve a common purpose, which is to provide anatomically defined landmarks for structural and functional neuroimaging experiments. We review some of these applications in this section and in section “Spatial Topography of Basal Forebrain Vulnerability to Neurodegeneration.”

Several different research groups have used resting-state functional MRI data to validate the anatomical parcellation of the BF Ch compartments and evaluate the topographical organization of their projections (Figs. 10.2 and 10.3). These studies capitalize on spontaneous fluctuations in hemodynamic responses measured from individual voxels, the smallest unit of measurement in MRI brain volumes. By measuring these fluctuations at each voxel in the brain over a sequence of several hundred timepoints acquired every ~ 2 s, one can determine whether fluctuations in different voxels correlate with one another over time (Li et al., 2014; Kline et al., 2016; Teipel et al., 2017; Zhang et al., 2017; Markello et al., 2018; Fritz et al., 2019; Yuan et al., 2019). These voxel-wise correlations can be used to determine whether brain regions exhibit spatial clustering of correlated spontaneous fluctuations, i.e., different groups or parcels of neighboring voxels with shared connectivity profiles.

Yuan et al. (2019) used high-resolution 7 T functional MRI to parcel the entire Ch1–Ch4 BF volume based on clusters of voxels exhibiting correlated spontaneous fluctuations in resting-state hemodynamic responses. Consistent with a topographic organization, voxels within the BF masking area grouped into three distinct clusters based on the strength of their correlated hemodynamic fluctuations (Fig. 10.4A). These data-driven functional parcels combined areas Ch1, Ch2, and Ch3 mask all together into Cluster 1, the anterior part of the Ch4 masks into Cluster 2, and the posterior Ch4p into Cluster 3. These three clusters exhibited both overlapping and distinct functional connectivity profiles with different resting-state networks (Fig. 10.4B). For instance, all clusters showed connectivity with salience networks. Cluster 1 was significantly associated with the default mode network, posterior default mode, salience network, auditory and language networks, as well as premotor (precentral) network. Cluster 2 was significantly connected with the salience, attention, language, and

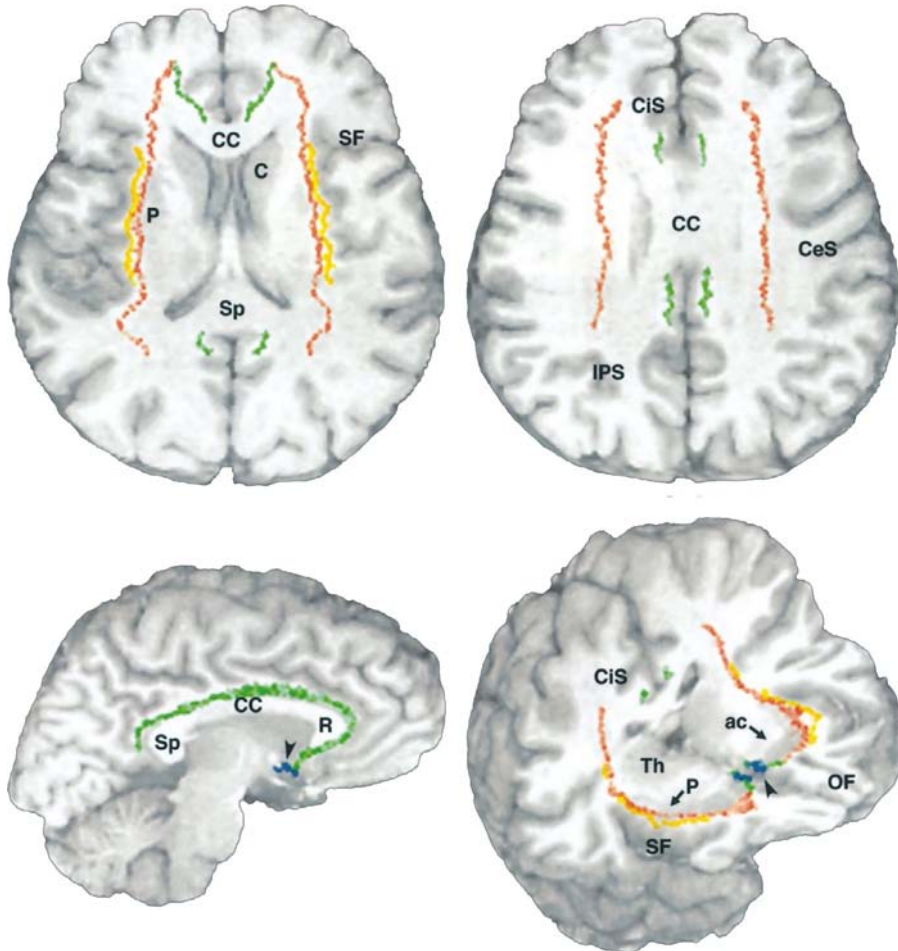


Fig. 10.3. Acetylcholinesterase (AChE) histochemistry was used to identify AChE-rich white matter fibers originating from the nucleus basalis of Meynert (equivalent to area Ch4 in Fig. 10.2) in five postmortem human brains. The locations of these projections in anatomical sections were traced onto a single-subject T1-weighted MRI volume. Cholinergic fiber bundles originating from the Ch4 region (*blue*) formed a medial pathway traveling within the cingulum (*green*) and a lateral pathway which further subdivided into a capsular component traveling within the external capsule and uncinate fasciculus (*red*), and a perisylvian component traveling within the claustrum (*orange*). The arrowhead points to the Ch4 neurons which give rise to the corticopetal cholinergic pathways. *ac*, anterior commissure; *C*, caudate; *CC*, corpus callosum; *CeS*, central sulcus; *CiS*, cingulate sulcus; *IPS*, intraparietal sulcus; *OF*, orbitofrontal cortex; *P*, putamen; *R*, rostrum of corpus callosum; *SF*, Sylvian fissure; *Sp*, splenium of corpus callosum; *Th*, thalamus. Adapted from Selden, N.R., Gitelman, D.R., Mesulam, M.-M., 1998. Trajectories of corticopetal cholinergic pathways within the cerebral hemispheres of the human brain *NeuroImage*, 7, S26.

premotor networks. Cluster 3 was preferentially correlated with the sensory-motor, premotor, attention, salience, and auditory networks.

Using independent resting-state fMRI datasets, [Fritz et al. \(2019\)](#) recovered highly similar three-cluster parcellations and connectivity maps for the BF. Taken together, the resting-state parcellations of the BF indicate that correlated fluctuations in hemodynamic response can provide reliable estimates of subregional BF connectivity.

In vivo structural MRI can also be used to measure the brain's white matter microstructure. In particular, diffusion tensor imaging measures the rate of water diffusion along axon bundles in the white matter. Diffusion tensor

tractography can then be used to infer the principal direction of this water diffusion and thus the white matter connectivity of “streamlines” between distal regions of the brain. Tractography has been used to evaluate the topography of white matter projections in the BF nucleus basalis of Meynert (Ch4) in living humans ([Liu et al., 2017](#); [Nemy et al., 2020](#)), guided by the postmortem work of [Selden et al. \(1998\)](#). These two in vivo imaging studies recovered medial and lateral white matter Ch4 pathways that largely resemble those identified postmortem by [Selden et al. \(1998\)](#), indicating that diffusion tractography is a promising tool for further evaluating the BF projectome. A complete mapping of the human

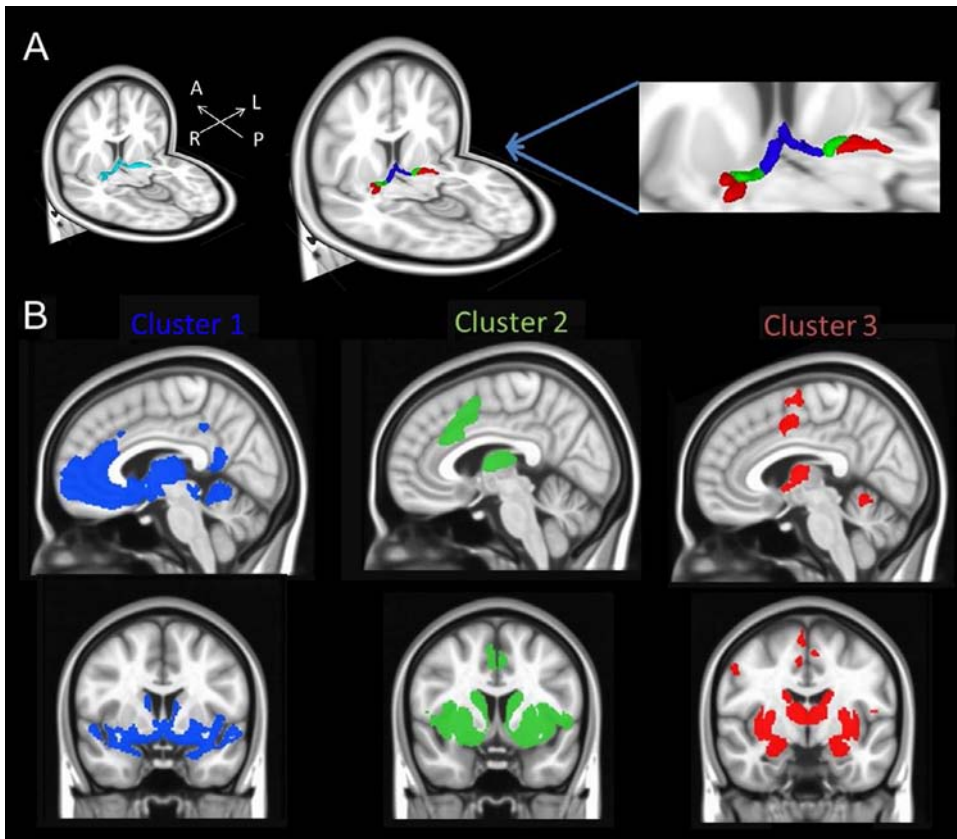


Fig. 10.4. Parcellation of the human cholinergic BF and its projections from 7 T resting-state fMRI. (A) *Left:* The light blue region of interest corresponds to the anatomical BF atlas (Fig. 10.2). *Right:* Voxels colocalized with the basal forebrain (BF) cholinergic (Ch) compartments grouped into three distinct clusters based on the strength of their correlated hemodynamic fluctuations. These data-driven functional parcels combined areas Ch1, Ch2, and Ch3 masks all together into Cluster 1, the anterior part of the Ch4 mask into Cluster 2, and the posterior Ch4p into Cluster 3. (B) These three clusters exhibited both overlapping and distinct functional connectivity profiles with other brain areas. Adapted from Yuan, R., Biswal, B.B., Zaborszky, L., 2019. Functional subdivisions of magnocellular cell groups in human basal forebrain: test-retest resting-state study at ultra-high field, and meta-analysis. *Cerebral Cortex Oxford Academic*, 29, 2844–2858.

BF projectome, integrating ex vivo and in vivo datasets with coverage of the entire set of Ch compartments (Ch1-4p) is lacking.

Because in vivo MRI techniques such as resting-state functional MRI and diffusion tensor imaging lack cell-type specificity, the ability to differentiate BF cholinergic compartments and their projections from other intermingled neurons represents a major methodological challenge. For example, we cannot infer from standard T1-weighted MRI assays of bulk gray and white matter tissue whether the tissue localized by the BF Ch compartments are specific to cholinergic cells or some combination of other cell types known to copopulate this region (Brashear et al., 1986; Freund and Gulyás, 1991; Freund and Meskenaite, 1992; Gritti et al., 1993, 2003, 2006; Hur and Zaborszky, 2005; Henny and Jones, 2008; Lin et al., 2015). Moreover, the interpretation of BF functional and structural connectivity patterns is complicated

by the fact that these indices likely reflect both mono-synaptic and polysynaptic anterograde and retrograde connections (Skudlarski et al., 2008; Honey et al., 2009).

In vivo PET of the human basal forebrain compartments and projections

Positron emission tomography (PET) is emerging as a promising tool for cell-type specific imaging of cholinergic projections in the living human brain. There are multiple PET radiotracers for imaging the cholinergic system, including radiotracers which bind to nicotinic (Horti et al., 2010) and muscarinic receptors (Malmquist et al., 2018), as well as the enzyme acetylcholinesterase (Kikuchi et al., 2013; Richter et al., 2018, 2019). A full review of these postsynaptic radiotracers and their ongoing development for clinical use is beyond the scope of this chapter. Here, we focus on a well-established PET radiotracer for imaging the presynaptic cholinergic system: ^{18}F -fluoroethoxybenzovesamicol

(^{18}F) FEOBV) (Mulholland et al., 1998); see also Tu et al. (2015) for a chemically similar radiotracer (^{18}F) VAT).

The [^{18}F] FEOBV radiotracer binds with high sensitivity and specificity to the vesicular acetylcholine transporter (VAcHT), a glycoprotein which is expressed in all presynaptic cholinergic nerve terminals of the mammalian central nervous system. Consistent with a topographic organization of cholinergic projections, experimentally induced lesions of cholinergic neurons in rats selectively reduces [^{18}F] FEOBV binding in the subcortical and cortical areas receiving their projections (Parent et al., 2012; Cyr et al., 2014). More recently, a first-in-human in vivo [^{18}F] FEOBV PET study (Aghourian et al., 2017) provided a whole brain cell-type specific map of brain areas that receive cholinergic input. This study also demonstrated that [^{18}F] FEOBV was decreased in the brains of cognitively impaired patients with probable AD (confirmed by a separate PET scan of amyloid binding). Further work in cognitively normal young and older adult humans has shown that [^{18}F] FEOBV PET is sensitive to age-related changes in central cholinergic integrity (Albin et al., 2018). In these studies, the observed spatial distribution of [^{18}F] FEOBV binding throughout the brain closely mirrors the expected spatial distribution of the cholinergic projection zones based on prior anatomical studies (Fig. 10.5). Collectively, these findings suggest that PET imaging of presynaptic VAcHT expression might serve as a powerful complementary approach to MRI methods for basic neuroscience research examining the topographical organization of the BF cholinergic system.

SPATIAL TOPOGRAPHY OF BASAL FOREBRAIN VULNERABILITY TO NEURODEGENERATION

The development of digital atlases for the BF cholinergic space (Teipel et al., 2005; Zaborszky et al., 2008; Kilimann et al., 2014) opened the way to investigate how in vivo structural changes in BF compartments correlate with topography of cortical pathology and progression of cognitive symptoms in neurodegenerative diseases of aging. In combination with the availability of large multimodal longitudinal neuroimaging datasets, such as the Alzheimer's Disease Neuroimaging Initiative (Mueller et al., 2005) and Parkinson's progressive markers initiative (Schrug et al., 2017), there has been a resurgence of interest in and evidence for the primacy of the cholinergic lesion in age-related neurodegenerative disease such as AD, and more recently, in PD. Later we review some of the work over the past 10 years which has combined probabilistic mapping of the human BF with in vivo neuroimaging, neuropsychological, and biospecimen data to examine subregional neurodegeneration of this structure in AD and PD.

Topography of basal forebrain vulnerability in Alzheimer's disease

Several cross-sectional structural MRI studies evaluating gray matter volume within the predefined BF atlas (e.g., Fig. 10.6) have consistently found evidence that subregional degeneration is not uniformly distributed across the Ch compartments in clinically diagnosed MCI and AD patients compared to age-matched controls (Teipel et al., 2005, 2011; Grothe et al., 2010, 2012; Kilimann et al., 2014; Cantero et al., 2017, 2020). These studies show that the posterolateral Ch4 and especially Ch4p BF compartments exhibit larger magnitudes of gray matter loss in MCI-AD compared to the more anteromedial Ch1–3 compartments (Fig. 10.6). By incorporating cerebrospinal fluid biomarkers of amyloid and tau as staging criteria, more recent cross-sectional studies have demonstrated the caudo-rostral axis of BF vulnerability at even earlier “preclinical” stages of AD, e.g., in cognitively normal adults with abnormal cerebrospinal fluid biomarkers of tau pathology (Cantero et al., 2020).

Studies which leverage both longitudinal structural MRI and cerebrospinal fluid biomarkers of amyloid-beta and tau neuropathology as staging criteria have enabled still further sensitivity to the cholinergic BF lesion in AD (Schmitz and Spreng, 2016; Schmitz et al., 2018, 2020; Fernandez-Cabello et al., 2020). These studies showed that annual rates of gray matter loss in the Ch4 and Chp BF subregion are detectable in cognitively normal older adults with abnormal cerebrospinal fluid biomarkers of amyloid and tau pathology. Moreover, degeneration of Ch4/Ch4p was consistently found to both precede and predict longitudinal degeneration in other vulnerable brain areas associated with the earliest phases of AD, such as the entorhinal, transentorhinal and perirhinal cortices. For instance, Fernandez-Cabello et al., 2020 showed that the Ch4/Ch4p→entorhinal predictive staging pattern of neurodegenerative spread is replicable between two large independent samples of older adults tracked over 2 years with longitudinal MRI. The strength of this pattern was significantly increased among older adults with abnormal CSF amyloid-beta and tau markers of neuropathology (Fig. 10.7A).

Follow-up exploratory whole brain staging models (not restricted to the entorhinal cortical region of interest) revealed that the medial temporal lobe structure most strongly expressing the predictive relationship from BF Ch4/Ch4p was in the adjacent transentorhinal region (Fig. 10.7B). These findings map well with known routes of BF cholinergic projections in rodent models, which show that the entorhinal cortex receives projections from the rostral Ch compartment, and the perirhinal cortex (Brodmann area 35) receives cholinergic projections from the caudal Ch compartment (Kondo and Zaborszky,

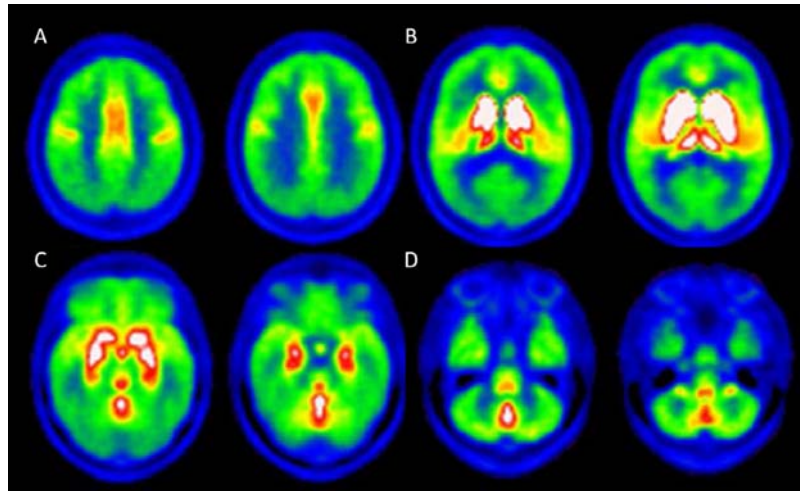


Fig. 10.5. PET imaging of the cholinergic projection system with [^{18}F] FEOBV (average of 13 young adults, mean age: 25 years, range: 20–38 years). The radiotracer binds to the vesicular acetylcholine transporter (VAcHT) glycoprotein, which is expressed selectively on the presynaptic nerve terminals of intrinsic cholinergic neurons. (A and B) Axial slices showing higher [^{18}F] FEOBV PET binding (warmer colors; white maximum) in units of standard uptake value ratio (SUVR) relative to the white matter. Each panel shows two closely situated slices in descending order (superior to inferior). (A) Neocortical binding sites in anterior cingulate and supplementary motor areas. (B) Dorsal striatal binding sites in the caudate nucleus and putamen, extending to the globus pallidus and thalamus. (C) Ventral striatal and pallidal binding sites (nucleus accumbens and basal forebrain), amygdala, hippocampus, and entorhinal cortex. (D) Mesopontine and cerebellar binding sites in tegmental nuclei, vermis, and flocculus. Figure adapted from Albin, R.L., Bohnen, N.I., Muller, M.L., et al., 2018. Regional vesicular acetylcholine transporter distribution in human brain: a [^{18}F] fluoroethoxybenzovesamicol positron emission tomography study. *J Comp Neurol Wiley Online Library*, 526, 2884–2897.

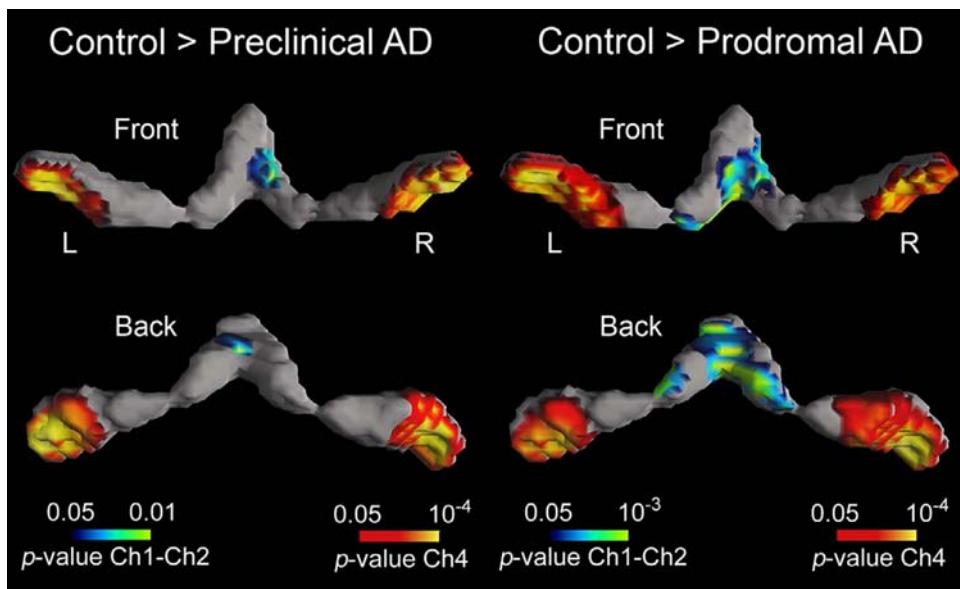


Fig. 10.6. Subregional degeneration of the BF in cognitively normal neurotypical older adults compared to preclinical adults (*left side*) and prodromal adults (*right side*). Preclinical status was defined as cognitively normal with abnormal cerebrospinal fluid biomarkers of amyloid and tau. Prodromal status was defined as mild cognitive impairment with abnormal cerebrospinal fluid biomarkers of amyloid and tau. In earlier (preclinical) stages of AD, subregional BF degeneration is stronger in posterolateral Ch4/Ch4p compartments (*warm clusters*) compared to anteromedial Ch1–3 compartments (*cool clusters*). AD, Alzheimer's disease; Ch, cholinergic cells. Adapted from Cantero, J.L., Atienza, M., Lage, C., et al., 2020. Atrophy of basal forebrain initiates with tau pathology in individuals at risk for Alzheimer's disease. *Cereb Cortex* 30, 2083–2098.

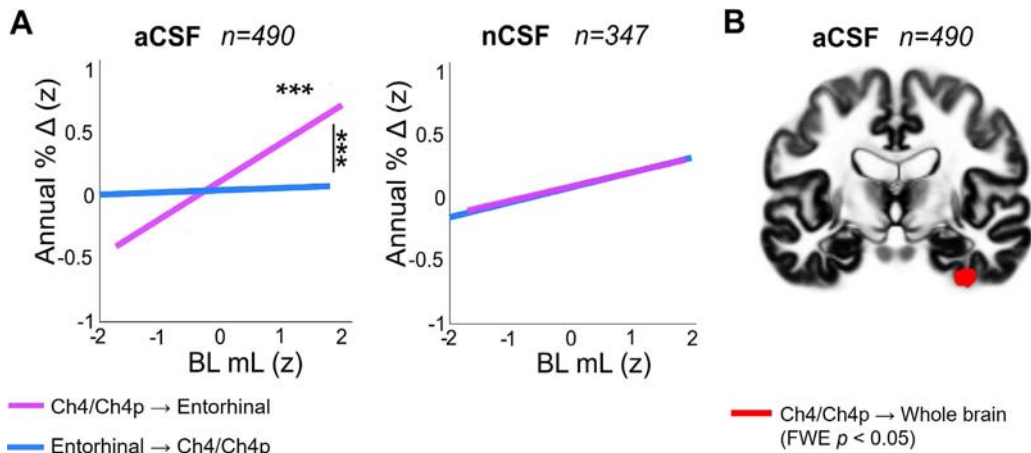


Fig. 10.7. Baseline gray matter volume (x -axes: BL mL = baseline milliliters) predicting subsequent longitudinal change in gray matter volume (y -axes: annual percent change in gray matter volume over 2 years, follow-up—baseline). (A) A priori region of interest analyses in BF Ch4/4p and entorhinal cortex. (Left plot) Baseline BF Ch4/4p gray matter volume predicts longitudinal change in entorhinal gray matter volume (magenta) more strongly compared to the reverse model (blue). The Ch4/4p → Entorhinal relationship was stronger among older adults with abnormal cerebrospinal fluid biomarkers of amyloid and phosphorylated tau pathology (aCSF, $n = 490$ adults). (Right plot) No difference between models was detected in older adults with normal CSF levels of amyloid and phosphorylated tau pathology (nCSF, $n = 347$ adults). (B) Exploratory whole brain analysis. When examining where baseline BF Ch4/4p gray matter volume predicts longitudinal changes in whole brain gray matter volume, suprathreshold voxels are restricted to the transentorhinal cortex (family-wise error corrected $P < 0.05$). Annual % Δ , annualized percent change in gray matter volume; aCSF, abnormal cerebrospinal fluid ratio of amyloid-beta concentration over phosphorylated tau concentration; BL mL, baseline gray matter volume in milliliters; FWE, family-wise error corrected P -value; nCSF, normal cerebrospinal fluid ratio of amyloid-beta concentration over phosphorylated tau concentration. Adapted from Fernandez-Cabello, S., Kronbichler, M., Van Dijk, K.R.A., et al., 2020. Basal forebrain volume reliably predicts the cortical spread of Alzheimer’s degeneration. *Brain* 143, 993–1009.

2016). In humans, this corresponds to the transentorhinal area, through which, like a funnel, the various neocortical areas project to the hippocampus. Disruption of cholinergic transmission in the transentorhinal/perirhinal cortices impairs object recognition (Tang et al., 1997; Winters et al., 2004). The emergence of localized tau pathology in human postmortem histological samples isolates cortical Stage 1 in the Braak predictive pathological staging model of AD (Braak and Del Tredici, 2015). The findings of Fernandez-Cabello et al. (2020) indicate that loss of Ch4/4p structural integrity precedes and predicts transentorhinal/entorhinal cortical degeneration, though higher resolution structural MRI data will be necessary to more conclusively disambiguate patterns of gray matter loss in these closely situated areas.

As mentioned in section “Topographic Projections of the Basal Forebrain in Humans,” the use of MRI data to delineate neurobiologically grounded topographic relationships between the BF and cortex requires multiple assumptions. One major assumption is that the interregional relationships in bulk gray and white matter structure observed between BF compartments and other brain areas reflect cholinergic projections. Multimodal imaging, integrating structural MRI, and [^{18}F] FEOBV provides initial clues in support of this assumption

(Schmitz et al., 2018). Patterns of longitudinal gray matter loss in the brain, assessed from structural MRI, tend to overlap patterns of cholinergic denervation assessed from [^{18}F] FEOBV PET (Fig. 10.8), implying that some proportion of cortical degeneration might reflect spatial topographies of cholinergic denervation. More work is needed to examine the interplay and timing of cholinergic denervation with the molecular and structural neuroimaging markers of AD pathology.

Topography of basal forebrain vulnerability in Parkinson’s and dementia with Lewy bodies

Pronounced BF cholinergic degeneration is observed in PD patients with dementia (PDD) and in patients with Lewy body dementia (LBD). Alpha-synuclein proteinopathy, which is common to PD, PDD, and LBD (Spillantini et al., 1997), may therefore also contribute to BF cholinergic degeneration. However, like amyloid and tau proteinopathies in AD, the causal mechanisms of BF cholinergic degeneration remain to be determined. Later we review some of the emerging evidence of an early BF cholinergic lesion in PDD and LBD.

The “dual-syndrome” hypothesis of PD (Williams-Gray et al., 2009, 2013; Kehagia et al., 2013) proposes

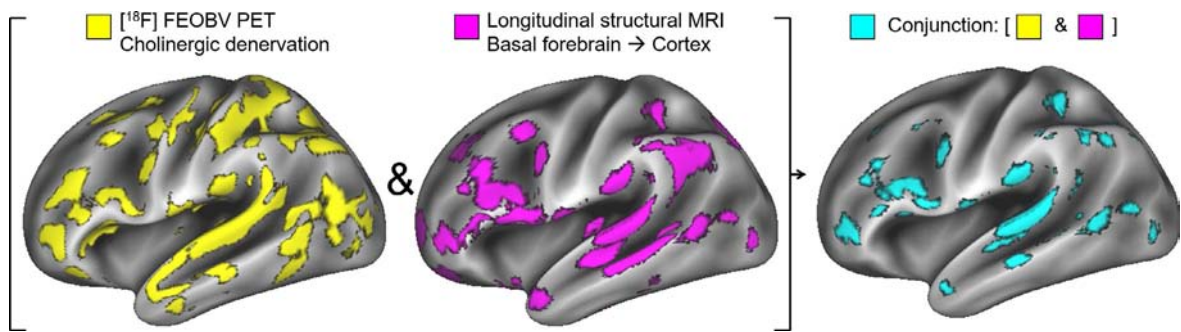


Fig. 10.8. From left to right, each cortical surface depicts a different measure of degeneration in the cholinergic systems of living older adult humans. (Left) Positron emission tomography (PET) was used in combination with the radiotracer [^{18}F] FEOBV, which binds to the cortical terminals of cholinergic neurons in the brain. Radiotracer binding intensity was reduced in older adults with early Alzheimer's disease (AD) compared to cognitively normal controls, indicating substantial loss of cholinergic terminals in the cortex (*yellow clusters*). (Middle) Magnetic resonance imaging (MRI) was used to measure changes in gray matter volume in the brain over a period of 2 years. Across individuals with early AD, loss of gray matter volume in the basal forebrain tracked with loss of gray matter volume in certain areas of the cortex (*purple clusters*). (Right) A “conjunction” map of the PET and MRI indices of cholinergic degeneration (left and middle panels, respectively) detected substantial overlap between these two distinct measures. Adapted from Schmitz, T.W., Mur, M., Aghourian, M., et al., 2018. Longitudinal Alzheimer's degeneration reflects the spatial topography of cholinergic basal forebrain projections. *Cell Rep* 24, 38–46.

that there are two phenotypes of PD with potentially distinct genetic contributions. The first PD phenotype is characterized by degeneration of dopaminergic neurons located in the substantia nigra pars compacta, the consequent loss of dopaminergic innervation in fronto-striatal circuits, and executive impairments. Patients exhibiting the dopaminergic PD phenotype respond well to dopaminergic agonists and typically do not progress to dementia. The second PD phenotype is characterized by memory and visuospatial impairments that rapidly progress to dementia, i.e., PDD. Patients with this neuropsychological profile respond well to the same classes of drugs used to treat AD, namely the acetylcholinesterase inhibitors, implying that PDD may represent a distinct cholinergic PD phenotype.

Following from the prediction that PDD should more strongly predict BF cholinergic degeneration than PD without dementia, Ray et al. (2018) used structural MRI to classify newly diagnosed cognitively normal PD patients into two groups based on MRI measures of their BF volume. This strategy produced two subgroups of PD patients, a control group and a group with abnormally small BF volume (one standard deviation below controls). They then tracked cognitive changes in these individuals over 5 years. Consistent with the dual-syndrome hypothesis, PD patients with smaller BF volumes on their baseline MRI scan showed greater decline on measures of recall memory and semantic fluency compared to control PD patients, implying greater temporoparietal degeneration. The groups declined at similar rates on measures of executive function. Other structural MRI studies have since shown that the

cholinergic PDD phenotype, like AD, preferentially impacts gray matter loss in the posterolateral Ch4/Ch4p BF compartments over the anteromedial C1/Ch2 compartments (Schulz et al., 2018; Pereira et al., 2020).

LBD is distinguished from PDD based on the clinical onset of cognitive symptoms relative to motor symptoms (cognitive precede motor deficits in LBD and vice versa in PDD). Nejad-Davarani et al. (2019) examined LBD patients with [^{18}F] FEOBV PET and observed evidence of a pronounced loss in cholinergic innervation throughout the brain. Due to the limited spatial resolution of PET, subregional BF analyses were not performed in that study. Nevertheless, these collective findings imply that AD, PDD, and LBD may share a common pathophysiological basis linked to early degeneration of the posterolateral BF cholinergic system.

Selective neuronal vulnerability to neurodegeneration: Factors and future directions

If the posterolateral BF cholinergic cell populations degenerate earlier than anteromedial populations in certain neurodegenerative diseases; we need to understand why this is so. The factors contributing to a local topography of degeneration in the BF may reveal important insights into cellular organization and vulnerability. The posterolateral and anteromedial cholinergic cell groups are thought to arise from distinct progenitors (Semba and Fibiger, 1988; Koh and Loy, 1989; De Carlos et al., 1995). Neurogenesis of the posterolateral cell populations occurs earlier in the developing rodent brain compared to

the anteromedial cell populations (Semba and Fibiger, 1988). Projections from the posterolateral cell populations into lateral and medial cortices also arrive earlier than the anteromedial projections into the hippocampus (De Carlos et al., 1995). In line with the possibility of distinct germinal sources, transcriptional (Pombero et al., 2011) and electrophysiological (Laszlovszky et al., 2020) profiling of mouse BF cholinergic neurons indicates that the posterolateral and anteromedial populations have distinct molecular properties and functional roles. Clues to posterolateral cholinergic BF vulnerability might therefore be gleaned from a deeper understanding of their ontogeny.

Moving from basic to clinical neuroscience, abnormal posterolateral BF cholinergic degeneration appears to identify three populations of cognitively normal at-risk individuals: (1) those with amyloid and tau pathology who progress to Alzheimer's dementia, (2) those with alpha-synuclein pathology who progress to Parkinson's dementia, and (3) possibly those with alpha-synuclein pathology who progress to Lewy body dementia. For the neurologist, this presents both challenges and opportunities. Current diagnostic criteria are usually based on *whether* there are cognitive symptoms in addition to motor symptoms (this differentiates AD from LBD and PD) or *when* cognitive symptoms started relative to the onset of motor symptoms (this differentiates LBD from PD). However, in all three cases, there is a long preclinical window of posterolateral BF cholinergic degeneration, which precedes the emergence of early cognitive symptoms. After the emergence of early cognitive symptoms, continued degeneration of the posterolateral BF is closely related to the progression to dementia. Because clinical diagnoses are tethered to the onset of early to moderate cognitive symptoms in AD and LBD, preclinical stages of these two neurodegenerative diseases, possibly spanning up to a decade, are currently beyond the reach of clinical treatment.

With the continued development of biomarkers sensitive to BF cholinergic integrity, future translational research bridging basic and clinical neuroscience is poised to facilitate dramatic improvements in preclinical identification of at-risk AD and LBD patients, as well as identification of de novo PD patients at-risk for dementia. In turn, drugs aimed at protecting the vulnerabilities of posterolateral BF cholinergic neurons in the preclinical window represent a highly promising strategy for preventing dementia in at-risk older adults.

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